

## **RECORDS OF PHARMACEUTICAL AND BIOMEDICAL SCIENCES**



### Evaluation of the Possible Protective Effect of Vitamin E on Nicotine-Induced Nephrotoxicity in Rats

Walaa I. Mohammed<sup>a</sup>,\*, Ghada M. Ahmed<sup>b</sup>, Wafaa A. Abdellah<sup>a</sup>

<sup>a</sup>Department of Pharmacology, Faculty of Medicine, Sohag University, Egypt. <sup>b</sup>Department of Histology, Faculty of Medicine, Sohag University, Egypt

Abstract

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\*Correspondence Author: Fax: +20934600526 E-mail address: walaa\_mohamed@med.sohag.edu.eg The goal of the current work was to investigate whether vitamin E had any protective effects against nicotine-induced nephrotoxicity in rats. Thirty mature male rats, weighing 200-250 g, were placed into 3 groups at random. Rats in the control group obtained saline, whereas Nicotine group: rats supplemented with nicotine (0.8 mg/kg/day; s.c.). Vitamin E+ nicotine group: rats were co-administered vitamin E (100 mg/kg/day; orally) followed by nicotine. The experiment duration was 30 days. Exposure of rats to nicotine resulted in significant rise in serum urea and creatinine levels, serum total oxidant status, tumor necrosis Factor- $\alpha$  and kidney tissue nitric oxide, malondialdehyde and caspase-3 levels. Otherwise, significant decrease in total antioxidant capacity. Moreover, the kidney histopathology investigation revealed significant damage. In addition to significant decrease in rats' weight and increase in relative kidney weight. On the other hand, the supplementation of Vitamin E altered the biochemical parameters and the histopathological findings. The results demonstrated that supplementation of Vitamin E protects against nicotine- induced kidney oxidative stress, inflammation and apoptosis.

**Keywords:** Nicotine; vitamin E; nephrotoxicity; anti-oxidant; tumor necrosis factor- $\alpha$ ; caspase-3.

### **1. Introduction**

There is mounting evidence that smoking use seriously harm renal health and is among the factors contributing to chronic renal disease (CRD). The most prevalent harmful alkaloid in cigarette smoke is nicotine, which is mostly accountable for renal impairment (**Arany et al., 2011**). After intake, nicotine reaches the blood and briskly redistributes throughout the body to different organs. Almost all nicotine is metabolized in the liver with the production of cotinine which is the primary metabolite of nicotine and used as a sign of its intake. The kidney is likely in charge of eliminating the cotinine that the liver produces (**Sobkowiak and Lesicki, 2013**). Nicotine raises oxidative stress level and promotes the formation of oxygen free radicals and lipid peroxidation responsible for the harmful effect on the kidneys (Arany et al., 2016). Moreover, nicotine triggers the production of proinflammatory proteins e.g., nuclear factor kappa (NF- $\kappa$ B), which causes an inflammatory response. In renal tubular epithelial cells, nicotine produced apoptosis via causing the production of ROS and activation of the NF- $\kappa$ B signaling pathways. Furthermore, nicotine can increase urine osmolality and decrease free water clearance by directly promoting the release of antidiuretic hormone (ADH). Also, persisting exposure to nicotine decreased the glomerular filtration rate (GFR). Additionally, nicotine has proangiogenic potential,

which causes various types of glomerular damage (Sudheer et al., 2008; Kim et al., 2016).

Vitamin E (lipid soluble antioxidant) has the power to stop or postpone the onset of chronic diseases brought on by free radicals by preventing lipid peroxidation and oxidative DNA damage in the cells. **Iranloye and Oludare (2011)** investigated that vitamin E boosted the antioxidant levels in the liver and heart in nicotine-treated rats and lowered the high levels of biochemical oxidative biomarkers.

So, this experiment was done to investigate the underlying mechanisms of nicotine- induced nephrotoxicity and to assess the protective effect of vitamin E on nicotine- induced nephrotoxicity in rats.

### 2. Materials and Methods

### 2.1. Animals

Mature male rats (average weight 200 –250 gm) were utilized in the experiment. They were acquired from animal house Faculty of Medicine, Sohag University. Animals were kept at 22-24°C room temperature, in the normal light/dark cycle and were fed a common animal nutriment with free water access. The Institutional Animal Care and Use Committee (IACUC) Sohag University, Egypt authorised the experimental protocol (Sohag-IACUC approval protocol No.5/5/2023/01).

### 2.2. Drugs and chemicals

Vitamin E and Nicotine were utilized in the present study. They were acquired from Pharma Co. for pharmaceuticals and chemical industries, Egypt and Sigma Aldrich Company, England, respectively. Kits for measurement of kidney function tests gained from Egyptian Company for Biotechnology, Cairo, Egypt. Kits for measurement of total oxidant status (TOS), total antioxidant capacity (TAC), nitric oxide (NO) and malondialdehyde (MDA) levels were obtained from Biodiagnostic Company Pharmaceutical Industries, Egypt. Moreover, tumor necrosis Factor-alpha (TNF- $\alpha$ ) and caspase-3 kits were obtained from Wuhan EIAab Science Co. Ltd (China) and Elabscience Co, Egypt, respectively.

### 2.3. Experimental design

Experimental animals from all groups were left to acclimatize for one week, and then were distributed into three groups of ten animals each. The experiment duration was 30 days.

**Control group:** rats received 0.5ml saline subcutaneously (s.c)

**Nicotine group:** rats received nicotine; 0.8 mg/kg/day dissolved in saline s.c. (Azab et al., 2022).

**Vitamin E+nicotine group:** rats received vitamin E; 100 mg/kg/day orally (**Oyeyemi et al., 2015**) followed by nicotine; 0.8 mg/kg/day dissolved in saline s.c.

All the animals were weighed after the completion of the experiment. The serum from blood samples was separated by centrifugation and was utilized for the measurement of kidney function tests, TOS, TAC and TNF- $\alpha$ . Each rat's two kidneys were removed and weighed; the right kidney was employed to measure the NO, MDA and caspase-3 levels. While the left kidney was used for histological examination.

### 2.4. Determination of relative kidney weight

Relative kidney weight (RKW)  $\% = (Rt kidney weight+Lf kidney weight)/(Final body weight) \times 100$  (**Imafidon et al., 2016**).

#### 2.5. Assay of kidney function tests

Spectrophotometric measurements of serum urea and creatinine levels were performed using commercially available kits bought from the Egyptian Company for Biotechnology in Cairo, Egypt.

# **2.6.** Assay of total oxidant status & total antioxidant capacity

Serum level of TOS and TAC were determined by a colorimetric method according to **Fossati et al.** (1980) and **Koracevic et al.** (2001), respectively. Their absorbance was measured at 510 NM. TOS and TAC were expressed as mM/L.

### 2.7. Oxidative stress index calculation

As stated by **Harman** (1956), oxidative stress index (OSI) was calculated using the following formula:  $(TOS/TAC) \times 100$ .

#### 2.8. Assay of tumor necrosis Factor-alpha

Enzyme-linked immunosorbent assay (ELISA) ratspecific kit was used for measurement of serum TNF- $\alpha$ , and it was expressed as pg/ ml.

## 2.9. Assay of nitric oxide and malondialdehyde

Levels of NO and MDA in kidney tissue were measured by a colorimetric method in kidney tissue homogenate according to **Montgomery and Dymock (1961)** and **Ohkawa et al. (1979)**. respectively. The sample's nitrite level was expressed as  $\mu$ mol/g tissue. While MDA level was expressed as nmol/g tissue.

### 2.10. Assay of caspase 3

Kidney tissue caspase-3 was measured using ELISA kit. Caspase 3 level was expressed as ng/ g tissue.

### 2.11. Histopathological examination

Following fixation in 10% formal saline, parts of the right kidney tissues collected and dehydrated in ascending grades of alcohol concentrations, and then embedded in Paraffin wax. After that, light microscope examination of 5 m thick sections stained with hematoxylin and eosin (H&E) was performed (**Bancroft and Gamble, 2002**).

### 2.12. Statistical analysis of data

Mean  $\pm$  SE was used to express the values. Tukey post hoc analysis was carried out after the one-way analysis of variance test (ANOVA) to determine whether there were any differences between the groups. At *p*<0.05, differences were considered significant. SPSS program, software package (version 26) was used in the analysis.

### 3. Results

# **3.1.** Effect of vitamin E on body weight and relative kidney weight

Nicotine administration produced significant (p<0.05) decrease in rats' body weight compared to the control group. Moreover, vitamin E administration produced significant (p<0.05) increase in rats' body weight compared to nicotine group (**Table 1**). As regard RKW nicotine group displayed a significant (p<0.05) elevation in comparison to the control group. However, vitamin E + nicotine group showed significant (p<0.05) decrease compared to nicotine group (**Table 1**).

### 3.2. Effect of vitamin E on kidney function

As shown in **Table 2**, serum urea and creatinine showed significant (p<0.05) rise in nicotine group in comparison to the control group. Whereas, vitamin E + nicotine group showed significant (p<0.05) diminution in serum urea and creatinine compared to nicotine group.

## **3.3.** Effect of vitamin E on total oxidant status & total antioxidant capacity

Total oxidant status in nicotine group raised significantly (p < 0.05) in comparison to the control group. However, vitamin E + nicotine group produced significant dwindling (p < 0.05) in serum TOS in comparison to the nicotine group (**Table 3**).

Parameter	Control	Nicotine	Vitamin E + Nicotine
Body weight (g)	$232\pm5.33$	$186^* \pm 4.52$	228•±11.23
Kidney weight (g)	$1.01\pm0.04$	$1.62^{*} \pm 0.09$	$1.12^{\bullet} \pm 0.07$
RKW	$0.44\pm0.02$	$0.87^{\ast}\pm0.05$	$0.50^{\bullet} \pm 0.04$

Table 1. Body weight, kidney weight and RKW of different studied groups

Mean  $\pm$  SE of 10 observations. RKW= Relative kidney weight. \* p < 0.05 vs the control group. • p < 0.05 vs the nicotine group.

#### Table 2. Kidney function tests of different studied groups

Parameter	Control	Nicotine	Vitamin E + Nicotine
Urea (mg/dl)	$42.9 \pm 1.89$	$55.9^{*} \pm 2.09$	44.7•±1.21
Creatinine (mg/dl)	$0.90\pm0.04$	$2.08^*\pm0.07$	$0.98^{\bullet} \pm 0.06$

Mean  $\pm$  SE of 10 observations. \* p < 0.05 vs the control group. • p < 0.05 vs the nicotine group.

Table 3. Serum TOS, TAC and OSI of different studied groups

Parameter	Control	Nicotine	Vitamin E + Nicotine	
TOS (mM/L)	$0.335 \pm 0.029$	$0.736^{*} \pm 4.52$	0.382• ± 0.024	
TAC (mM/L)	$2.62\pm0.195$	$0.9^{*} \pm 0.043$	$2.03^{\bullet} \pm 0.105$	
OSI	$13.77 \pm 1.89$	84.12* ± 6.53	19.03•±1.29	

Mean  $\pm$  SE of 10 observations. TOS= Total oxidant status, TAC= Total antioxidant capacity, OSI= Oxidative stress index. \* p < 0.05 vs the control group. • p < 0.05 vs the nicotine group.

Total antioxidant capacity in nicotine group was significantly (p<0.05) decreases compared to the control group. Whereas, in vitamin E+nicotine group it exhibited a significant (p<0.05) elevation compared to nicotine group (**Table 3**).

## **3.4.** Effect of vitamin E on oxidative stress index

**Table 3** showed significant (p < 0.05) increase in OSI in nicotine group in comparison to the control group. Treatment of the rats with vitamin E + nicotine produced significant (p < 0.05) decrease in OSI compared to in nicotine group.

# 3.5. Effect of vitamin E on tumor necrosis Factor- $\alpha$

Tumor necrosis Factor- $\alpha$  level exhibited a significant (p<0.05) elevation in nicotine group in comparison to the control group. On the other hand, vitamin E + nicotine group exhibited a significant (p<0.05) decrease in TNF- $\alpha$  compared to in nicotine group (**Fig. 1**).



Figure 1. Serum tumor necrosis factor- $\alpha$  of different studied groups. Mean $\pm$  SE of 10 observations. TNF- $\alpha$  = tumor necrosis factor- $\alpha$ .\* p< 0.05 vs the control group. • p< 0.05 vs the nicotine group.

#### 3.6. Effect of vitamin E on NO and MDA

Nicotine group revealed a significant (P<0.05) rise in kidney NO level in comparison to the control group. Whilst, vitamin E + nicotine group revealed a significant (p<0.05) reduction in kidney NO level in comparison to nicotine group (**Fig. 2**).

Malondialdehyde significantly (p<0.05) increased in nicotine group compared to the control group. While, it significantly (p<0.05) decreased in vitamin E + nicotine group compared to nicotine group (**Fig. 3**).



Figure 2. Kidney NO level of different studied groups. Mean $\pm$  SE of 10 observations. NO = nitric oxide. \* p < 0.05 vs the control group. • p < 0.05 vs the nicotine group.



Figure 3. Kidney MDA level of different studied groups. Mean $\pm$  SE of 10 observations. MDA = malondialdehyde. \* p < 0.05 vs the control group. • p < 0.05 vs the nicotine group.

### 3.7. Effect of vitamin E on caspase-3

As regard caspase-3 level, nicotine group expressed a significant (p<0.05) elevation in kidney caspase-3 level in comparison to the control group. Nevertheless, compared to the nicotine group, the vitamin E + nicotine group significantly (p<0.05) decreased kidney caspase-3 level (**Fig. 4**).



Figure 4. Kidney caspase-3 level of different studied groups. Mean $\pm$  SE of 10 observations. \* p < 0.05 vs the control group. • p < 0.05 vs the nicotine group.

### 3.8. Histopathological changes

Kidney section of control group showed the renal cortex contains the renal corpuscle and the renal tubules. The renal corpuscle is formed of renal glomeruli surrounded by the Bowman's capsule which is formed of parietal and visceral layers. The proximal tubules (P.T) are lined with cuboidal cells with deeply acidophilic cytoplasm and rounded vesicular nuclei. The apical cell membranes have brush borders. The distal tubules (D.T) exhibit wide lumen and are lined with low cuboidal cells with rounded vesicular nuclei with no brush borders (Fig 5A). Otherwise, kidney of nicotine group showed that, the renal corpuscle had dilated urinary space, congestion of the glomerular capillaries and Most of the proximal glomerular separation. tubules showed dilated lumen and degenerated brush borders. Some proximal and distal tubular cells had deeply stained pyknotic nuclei. Interstitial tissue showed inflammatory cell infiltration (Fig 5B). Examination of kidney sections of vitamin E+nicotine group showed an apparent decrease in the urinary space with less congested glomerular capillaries and glomerular separation compared to nicotine group. Some proximal and distal tubules restored their normal shape. Some proximal and distal tubular cells still had pyknotic nuclei (Fig 5C).

### 4. Discussion

Smoking is a significant risk factor that has the unique ability to make renal impairment worsened (**Orth and Hallan, 2008**). Both males and females

who smoke cigarettes are at an increased risk of developing chronic kidney disease or passing away from end-stage renal disease (Ishani et al., 2006). Smoking greatly accelerates the progression of renal disease by causing microvascular injury and hastening glomerulosclerosis. Smoking also seriously affects kidney structure and function (Ramalingam et al., 2019). Over 4000 different substances are found in typical cigarette smoke, including carbon monoxide, reactive oxygen species, ketones, and stable reactive aldehydes (Huang et al., 2005). Nicotine is one of the biologically stable and active substances found in tobacco that is easily absorbed through both active and passive smoking, and is responsible for the addictive effect of smoking. Moreover, it plays a significant role in the etiology of many diseases, including kidney disease. Nicotine exposure through chronic cigarette smoking appears to be the reason that hasten the microvascular complications. Furthermore, nicotine downregulates eNOS, which is a first step in the pathophysiology of kidney injury, and causes vascular endothelial dysfunction in rats. Additionally, smoking has been shown to nephropathy exacerbate by triggering hyperlipidemia and high-grade oxidative stress (Balakumar et al., 2009; Chakkarwar, 2011; Ben Saad et al., 2019; Zhang et al., 2019).

Accordingly, the current research was prepared to assess the underlying mechanisms of nicotineinduced nephrotoxicity. Moreover, to assess the protective effect of vitamin E on nicotine- induced nephrotoxicity in rats. It was observed in the present study that giving rats nicotine reduced their appetite, which most likely caused them to weigh less than controls. Contrary significant increase in RKW in nicotine group compared to the control group can be attributed to the reduction of body weight and increased kidneys weight which may results from edema of the kidneys due to nicotineinduced tubular necrosis. Moreover, nicotine administration produced significant rise in serum urea and creatinine levels and pathological changes in glomeruli, suggesting that renal impairment was brought on by nicotine exposure. Our results are in agreement with the results of Ibrahim et al. (2016) and Azab et al. (2022). According to Ahmed et al. (2015), smokers' urea and creatinine levels were considerably higher than those of the control group that might be due to increase renovascular resistance as a results of cigarette smoking. Increased renovascular resistance significantly decrease GFR, filtration fraction, and renal plasma blood. Moreover, the decrease in GFR will cause the distal tubular flow rate to drop, which will raise



**Figure 3. Kidney histopathological changes.** (A) Photomicrograph of the control rats showing; the renal corpuscle (RC), glomerular capillaries(G), urinary space (U), proximal tubule (P.T), and distal tubule (D.T), (B) the rats treated with nicotine showing renal corpuscle (RC), congestion of glomerular capillaries (G) with glomerular separation (thick arrow) and widening of urinary space (U). Some P.T and D.T showed pyknotic nuclei (curved arrow) with loss of brush border of P.T. Note inflammatory cell infiltration in the interstitial tissue (blue arrow), (C) the rats treated with nicotine and vitamin E showing renal corpuscle (RC) with less glomerular (G) separation and congestion with decrease of urinary space (U) compared to group II. Some proximal tubules (P.T) and distal tubules (D.T) still showed pyknotic nuclei (curved arrow). Interstitial tissue showed less inflammatory cell infiltration compared to group II. (H&E, A: x200, B and C:x400)

urea reabsorption (JoAnn and Robert, 2011). The kidneys easily filter creatinine, a non-protein waste product, and because the serum creatinine level depends on the GFR, it is regarded as a sign of renal dysfunction (Zhang et al., 2019).

Oxidative stress almost always related to organ toxicity, in the present study administration of nicotine produced significant increase in serum TOS and TNF- $\alpha$  levels, kidney tissue MDA, NO and caspase-3 levels with significant decrease in serum TAC compared to the control group. Moreover, it increases OSI compared to the control group. Numerous investigations have demonstrated

that nicotine has an oxidative stress potential and mitochondrial ROS release damages organs and tissues (Arany et al., 2011; Mohamed et al., 2022). Furthermore, Mohamed et al. (2022) reported that nicotine-induced oxidative damage together with its antioxidant suppression have been found to exacerbate tubular necrosis in rats' renal tissue and might be the cause of increased caspase-3 level in kidney tissue. The increased serum level of TNF- $\alpha$  by nicotine administration may be due to induction of proinflammatory cytokines and the overexpression of their genes. These cytokines can stimulate leukocytes to generate additional chemokines and pro-inflammatory cytokines, which will aggravate the inflammation and is the cause of inflammatory kidney state. TNF- $\alpha$  also activates the death receptor pathway, which leads to caspasemediated apoptosis (Sun et al., 2013). The present biochemical abnormalities are confirmed by histopathological abnormalities in the renal tissue in the form of dilated urinary space in renal corpuscle, congestion of the glomerular capillaries and glomerular separation. Most of the proximal tubules showed dilated lumen and degenerated brush borders. Some proximal and distal tubular cells had deeply stained pyknotic nuclei. And inflammatory cell infiltration in the interstitial tissue. These results are in accordance with the results of (Menshawy et al., 2019; Azab et al., 2022; Mohamed et al., 2022).

The present study revealed that concurrent treatment with vitamin E protected from nicotineinduced nephrotoxicity by improvement of body weight and RKW. In addition to significant decrease in serum urea and creatinine levels. Also, there were effective restoration of serum TOS, TAC and TNF- $\alpha$  levels. Moreover, kidney MDA, NO and caspase-3 levels were significantly decreased. The restoration of the renal architecture under a light microscope provides proof for this conclusion. The renal protective impacts of vitamin E might be allocated to its antioxidant and free radicals scavenging properties. These findings are in accordance with (Nematbakhsh et al., 2012; Abo-Elmaaty et al., 2020; Erdemli et al., 2020). Moreover, the anti-apoptotic activity of vitamin E was verified by a significant reduction of kidney tissue caspase-3 level. This is might be because vitamin E effects on the prevention of mitochondrial oxidative stress and the prevention of mitochondrial release of apoptotic signaling molecules into the cytoplasm (Weber et al., 2003). Furthermore, the radical scavenging ability of vitamin E may be the cause of its anti-inflammatory effects. Likewise, the anti-inflammatory effects of vitamin E could be explained by suppression of NFκB protein expression during the inflammatory process (Sarir et al., 2015).

### 5. Conclusion

The current study confirms nicotine-induced nephrotoxicity in rats based on the current biochemical findings, which are substantiated by histological data. This toxic effect is due to induction of oxidative stress, inflammatory mediators and apoptotic pathways. Treatment of the rats with vitamin E protects against this nephrotoxic effects.

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### **Conflicts of Interest**

The authors declare that there is no conflict of interest.

### **References:**

Abo-Elmaaty A.M., Behairy A., El-Naseery N.I., Abdel-Daim M.M., 2020. The protective efficacy of vitamin E and cod liver oil against cisplatininduced acute kidney injury in rats. Environmental Science and Pollution Research, 27, 44412-44426.

Ahmed M.M.E., Jawad A.S.A., Osman H.M., Shayoub M.E., 2015. The effect of smoking cigarette on kidney functions among sundaes peoples. International Journal of Development Research, 5(5), 4473-4475.

Arany I, Grifoni S, Clark J.S., Csongradi E, Maric C., Juncos L.A., 2011. Chronic nicotine exposure exacerbates acute renal ischemic injury. American Journal of Physiology-Renal Physiology, 301, 125-133.

Arany I., Hall S., Reed D.K., Reed C.T, Dixit M., 2016. Nicotine Enhances High-Fat Diet-Induced Oxidative Stress in the Kidney. Nicotine & Tobacco Research, 18, 1628–1634.

Azab A.E., Albasha M.O., Elnaif M.A., 2022. Renal Toxicity Induced by Nicotine in Male Albino Rats and Attenuation by Fenugreek Seeds and Curcumin. Biotechnology and Bioprocessing, 3(2), 2766-2314.

Balakumar P., Chakkarwar V.A., Krishan P., Singh M., 2009. Vascular endothelial dysfunction: a tug of war in diabetic nephropathy. Biomedicine & Pharmacotherapy, 63(3), 171-179.

Bancroft J.D., Gamble M., 2002. Theory and practice of histological techniques. Histopathology, Churchill Livingstone, Edinburgh, London, 5<sup>th</sup> edition.

Ben Saad A., Ncib S., Rjeibi I., Saidi I., Zouari N., 2019. Nephropro¬tective and antioxidant effect of green tea (Camellia sinensis) against nicotineinduced nephrotoxicity in rats and characteriza¬tion of its bioactive compounds by HPLC-DAD. Applied Physiology, Nutrition, and Metabolism, 44(11), 1134-1140. Chakkarwar V.A., 2011. Fenofibrate attenuates nicotine-induced vascular endothelial dysfunction in the rat. Vascular Pharmacology, 55(5-6), 163-168.

Erdemli M.E., Zayman E., Erdemli Z., Gul M., Gul S., Gozukara Bag H., 2020. Protective effects of melatonin and vitamin E in acetamiprid-induced nephrotoxicity. Environmental Science and Pollution Research, 27(9), 9202-9213.

Fossati P., Prencipe L., Berti G., 1980. Use of 3, 5dichloro-2-hydroxybenzenesulfonic acid/4aminophenazone chromogenic system in direct enzymic assay of uric acid in serum and urine. Clinical Chemistry, 26(2), 227-231.

Harman D., 1956. Aging: a theory based on free radical and radiation chemistry. Journals of Gerontology, 11(3), 298-300.

Huang M.F., Lin W.L., Ma Y.C., 2005. A study of reactive oxygen species in mainstream of cigarette. Indoor Air, 15, 135–140.

Ibrahim Z.S., Alkafafy M.E., Ahmed M.M., Soliman M.M., 2016. Renoprotective effect of curcumin against the combined oxidative stress of diabetes and nicotine in rats. Molecular Medicine Reports, 13(4), 3017-3026.

Imafidon C.E., Akomolafe R.O., Oladele A.A., 2016. Sexually dimorphic proteinuria in Wistar rats: relevance to clinical models. Pathophysiology 23, 51–59.

Iranloye B.O., Oludare G.O., 2011. Garlic and vitamin E provides antioxidant defense in tissues of female rats treated with nicotine. Nigerian Journal of Physiological Sciences, 26, 103–107.

Ishani A., Grandits G.A, Grimm R.H., Svendsen K.H., Collins A.J., Prineas R.J., Neaton J.D., 2006. Association of single measurements of dipstick proteinuria, estimated glomerular filtration rate, and hematocrit with 25-year incidence of end-stage renal disease in the multiple risk factor intervention trial. Clinical Journal of the American Society of Nephrology, 17, 1444–1452.

Lindenfeld, J., Schrier, R.W. 2011. Blood Urea Nitrogen A Marker for Adverse Effects of Loop Diuretics. Journal of the American College of Cardiology, 58(4), 383-385. Lee J., Kim S.W., 2016. Nicotine-Induced Apoptosis in Human Renal Proximal Tubular Epithelial Cells. PLoS ONE, 11, e0152591.

Koracevic D., Koracevic G., Djordjevic V., Andrejevic S., Cosic V., 2001. Method for the measurement of antioxidant activity in human fluids. Journal of Clinical Pathology, 54(5), 356-361.

Menshawy M.M., Sharaf W.M., Esmail R.S., Farrag A.R., 2019. Histopathological and histochemical effects of nicotine on the liver and kidney of adult male rats. Journal of The Arab Society for Medical Research, 14(1), 7.

Mohamed, A.A., El-Kholy, S.S., Dahran, N., El Bohy, K.M., Moustafa, G.G., Saber, T.M., Metwally, M.M.M., Gaber, R.A., Alqahtani, L.S., Mostafa-Hedeab, G., El-Shetry, E.S. 2022. Scrutinizing pathways of nicotine effect on renal Alpha-7 nicotinic acetylcholine receptor and Mitogen-activated protein kinase (MAPK) expression in Ehrlich ascites carcinoma-bearing mice: Role of Chlorella vulgaris. Gene, 837, 146697.

Montgomery H.A., Dymock J.F., 1961. The determination of nitrite in water. Analyst, 86, 414-416.

Nematbakhsh M., Ashrafi F., Safari T., Talebi A., Nasri H., Mortazavi M., Khazaei M., Baradaran-Mahdavi M.M., 2012. Administration of vitamin E and losartan as prophylaxes in cisplatin-induced nephrotoxicity model in rats. Journal of Nephrology 25(3), 410–417.

Ohkawa H., Ohishi N., Yagi K., 1979. Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. Analytical Biochemistry, 95 no. 2, 351–358.

Orth S.R., Hallan S.I., 2008. Smoking: A risk factor for progression of chronic kidney disease and for cardiovascular morbidity and mortality in renal patients–absence of evidence or evidence of absence. Clinical Journal of the American Society of Nephrology, 3, 226–236.

Oyeyemi W.A., Shittu S.T., Kolawole T.A., Ubanecheand P., Akinola A.O., 2015. Protective effect of vitamin E on nicotine induced reproductive toxicity in male rats. Nigerian Journal of Basic and Applied Sciences, 23(1),7-13. Ramalingam A., Santhanathas T., Shaukat Ali S., Zainala¬bidin S., 2019. Resveratrol Supplementation Protects Against Nic¬otine-Induced Kidney Injury. International Journal of Environmental Research and Public Health, 16(22), 4445.

Sarir H., Emdadifard G., Farhangfar H., TaheriChadorneshin H., 2015. Effect of vitamin E succinate on inflammatory cytokines induced by high-intensity interval training, Journal of Research in Medical Sciences, 20(12), 1177–1181.

Sobkowiak R., Lesicki A., 2013. Absorption, metabolism and excretion of nicotine in humans. Postepy. Biochemii, 59, 33-44.

Sudheer A. R., Muthukumaran S., Devipriya N., Devaraj H., Menon V. P., 2008. Influence of ferulic acid on nicotine-induced lipid peroxidation, DNA damage and inflammation in experimental rats as compared to N-acetylcysteine. Toxicology, 243, 317-329.

Sun Q., Meng Q.T., Jiang Y., Liu H.M., Lei S.Q., Su W.T., Duan W.N., Wu Y., Xia Z.Y., Xia Z.Y., 2013. Protective effect of ginsenoside Rb1 against intestinal ischemia-reperfusion induced acute renal injury in mice. PLoS ONE, 8, e80859.

Weber T., Dalen H., Andera L., Nègre-Salvayre A., Augé N., Sticha M., Plasilova M., 2003. Mitochondria play a central role in apoptosis induced by -tocopheryl succinate, and agent with antineoplastic activity: comparison with receptormediated pro-apoptotic signaling. Biochemistry, 42, 4277–4291.

Zhang Y., Chen Y., Zhang Y., Li P L., Li X., 2019. Contribution of cath¬epsin B-dependent Nlrp3 inflammasome activation to nico¬tine-induced endothelial barrier dysfunction. European Journal of Pharmacology, 865, 172-795.