

### **RECORDS OF PHARMACEUTICAL AND BIOMEDICAL SCIENCES**



# The emerging role of autophagy in the pathophysiology of diabetic neuropathy

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#### Abstract

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\*Correspondence Author: Tel: 0096551666602 E-mail address: ph.thekrayat@gmail.com Diabetic neuropathy (DN) is a common complication that is associated with diabetic patients. It affects almost half of diabetic patients worldwide. Peripheral DN is associated with neurodegeneration as evidenced by the degenerative loss of fibres in the peripheral nerves with concomitant decrease of nerve conduction Multiple mechanisms were proposed to have role in the pathophysiology of DN including oxidative stress, inflammation and recently autophagy. Autophagy is a cellular process that is present from yeast to mammals for the sake of recycle damaged organelles and protein in autophagosomes. Persistent hyperglycaemia causes an imbalance in the cellular autophagic pathways. Autophagy plays an important role in reserving the cellular homeostasis via its ability to recycle the damaged proteins and organelles. The link between the activation of autophagic flux with the control of the toxic effect of hyperglycaemia on the neuronal cells will be discussed in this review. The present review hypothesizes that upregulation of the autophagic pathways in neuronal cells may aid them to ameliorate the bioenergetic crisis and cellular damage concomitant with DN.

Keywords: Autophagy; Diabetic neuropathy; pathogenesis.

### **1. Introduction**

Long standing hyperglycemia associated with diabetic subjects can result in the development of various microvascular complications including diabetic neuropathy (DN). Symptoms of DN may disturb the quality of life of affected patients as they include tingling sensation in the extremities, burning allodynia and hyperalgesia pain, (Schreiber et al. 2015). Almost half of diabetic subjects of both types of diabetes suffer from DN (Zimmet 2009). Pregabalin and Duloxetine are among FDA approved drugs to control the neuropathic pain which is the main concern in patients suffering from DN. They act mainly by

interfering with the neurotransmitters involved in the pain processing pathways, thus they are capable to relieve symptoms associated with DN (Ziegler 2008). Several mechanisms are involved in the pathogenesis of DN including like oxidative/nitrosative stress, inflammation, endoplasmic reticulum stress, metabolic crisis and demyelination (Zenker et al. 2013). However, different drugs targeting any of these pathways fail to control the symptoms on large scale of clinical trials (Singh et al. 2014). This can be attributed to the involvement of more than one signaling pathway in the etiology of DN or to the specific stage of involvement of a certain pathway. Therefore, there is a need to search for drugs that are able to target

many of the involved pathways.

Autophagy is a cellular process that is present from veast to mammals for the sake of recycling damaged organelles and protein in autophagosomes. Autophagy is additionally involved in clearing cells undergone physiological programmed cell death (Mizushima 2007). Dysregulation of autophagy was reported in the pathogenesis of several diseases including cancer and neurodegenerative disorders (Ravikumar et al. 2010; Wong and Cuervo DN is 2010). Peripheral associated with neurodegeneration as evidenced by the degenerative loss of fibres in the peripheral nerves with concomitant decrease of nerve conduction (Reichling and Levine 2011). In the present review, the theoretical assumption of linking the activation of autophagic flux with the control of the toxic effect of hyperglycemia on the neuronal cells is discussed.

## 2. Regulation of Basic Machinery of Autophagy

Autophagy is a complex multistep cellular process that occurs with the aid of set of proteins named autophagy related proteins (Atg). The first step is the formation of a phagophore that is derived either from Golgi apparatus or endoplasmic reticulum (Mizushima et al. 2011). The phagophore is made by the combination of Atg 13 and Atg 17 (FIP200) with Atg1or Ulk 1 in mammals. Mammalian target of rapamycin (mTOR) kinase controls this step thus the autophagy is initiated upon the exposure to nutrient starvation and stress signals. In case of nutrient availability, mTOR can phosphorylate Atg13 thus prevents its combination with Atg1 and hence inhibits autophagy (Jung et al. 2010). The elongation and extension of phagophore is further controlled by the activity of class III PI3K enzyme produce phosphatidyl (Vps34)to inositol triphosphate (PI3P) from phosphatidyl inositol (PI). Beclin1 (mammalian homolog of Atg6) also helps to recruit other Atg proteins to the growing phagophore membrane (Glick et al. 2010).

The maturation involves the association of Atg5-Atg12 to Atg16L to form a complex attached to autophagosome. In addition, this Atg5-Atg12-Atg16L complex recruits microtubule associated protein light chain (LC3B) (mammalian homolog of Atg8) into the autophagosome (**Glick et al. 2010**). The matured autophagosome could be delivered to the lysosomes through endosomal trafficking that involves the activity of Rab7 GTPases and

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cytoskeletal motor proteins. The hydrolases of lysosomes can digest the engulfed cellular debris and deliver the basic subunits through permeases to cytosol for recycling (Longatti and Tooze 2009).

### **3.** Chaperone-mediated Autophagy in Diabetic Neuropathy

Chaperone-mediated autophagy (CMA) is an intracellular degradation process that involves the activity of lysosomes to breakdown the cytosolic protein (Kim and Koh 2021). Chaperones are mainly implicated in the repair of degraded proteins. Commonly neuropathic disorders the are characterised by the presence of intracellular protein aggregates (Xilouri et al. 2013). PMP22 is a key component of myelin sheath that was found to be aggregated in several neuropathies leading to the formation of aggresomes. Stimulation of autophagy contributes in the clearance of aggresomes thus the upregulation of chaperone receptors in the lysosomal membrane could be a useful therapeutic target to clear the protein aggregates (Ravikumar et al. 2009).

Diabetic neuropathy is characterized by the decrease in the nerve conduction and demyelination of either the motor or the sensory nerves or both. Enhanced activity of CMA could attenuate these symptoms (Mizushima et al. 2004). Autophagy can be induced by inhibition of mTORC that acts by phosphorylating the Atg13 and thus prevents its combination with Atg1 and inhibits the formation of autophagosomes (Jung et al. 2010). In addition, autophagy can be stimulated by another mechanism independent to mTOR involving the phosphoinositol and calcium signaling. Therefore, there are several drugs and natural products acting to activate autophagy either as mTOR inhibitors or inositol monophosphatase (IMPase) inhibitors were shown to protect against neuropathies.

**Figure 1** shows the postulation for the therapeutic effect of activators of autophagy on diabetic neuropathy. Examples for some drugs and their mode of action in protection against several models of neuropathies are presented in **Table 1**.

Regardless the signalling pathways involved in the activation of autophagy, it is expected that induction of autophagy can ameliorate the deleterious effects associated with DN. Autophagy induction can reduce the oxidative stress, enhance ATP production, and lessen apoptosis through BCL-2

activation (**Choi et al. 2013**). In addition, it can be postulated that drugs able to activate autophagy either by inhibiting mTOR or inositol monophosphatase can prevent the mitochondrial dysfunction and clear out the protein aggregates.

### 4. Modulation of autophagy in diabetic neuropathy

There is some evidence of accumulation of autophagosomes upon exposure of sera separated from type 2 diabetic neuropathic patients to neuroblastoma cells. This was confirmed by increased LC3-II immunoreactivity (**Towns et al. 2005**). In experimental model of DN, it was demonstrated that there was an impairment of autophagy reflux with concomitant mitochondrial dysfunction (**Yang et al. 2014**).

Melatonin is a potent antioxidant that was shown to reduce the neuronal excitability in a subset of the dorsal root ganglia (DRGs) neurons (**Tan et al. 2015, Oliveira-Abreu et al. 2018**). Intraperitoneal administration of melatonin alleviates oxaliplatininduced pain and neuropathic impairments in rats through boosting the autophagy pathway in peripheral neurons and DRG (**Areti et al. 2017**).

Progranulin is a trophic factor that supports neurons by suppressing microglia activation. It was found in neurons and microglia in the nervous system (Tanaka et al. 2013). Progranulin overexpression in DRG nociceptive neurons improves autophagy and pain behaviour in damaged nerves (Altmann et al. 2016). Another example is Chloroquine (CQ) that the lysosomal can inhibit proteases and autophagosome-lysosomal fusion processes, making it an autophagy blocker (Geng et al. 2010). Therefore, intrathecal injection of CO in naive mice accumulation of microtubulecauses spinal associated protein 1 light chain 3 (LC3) and p62, as well as significant mechanical hypersensitivity, implying a block in autophagosome clearance and demonstrating the role of the autophagic process in spinal pain processing mechanisms (Berliocchi et al. 2015).

### 5. Conclusion

Autophagy is an important pathway that involves the activity of lysosomal enzymes for the sake of recycle damaged proteins. In diabetic neuropathy, autophagy is crucial to recycle the damaged protein aggregates and lessen the mitochondrial damage. Thus, autophagy induction can enhance the survival chance in neurons and glia exposed to hyperglycemic conditions. Being a catabolic autophagy induction improves process, the bioenergetic status. Therefore, drugs especially natural ones that are able to induce autophagy could be tested as therapeutic tool to treat neuropathy.



Figure 1: Proposed theory for potential effect of activators of autophagy on diabetic neuropathy (Yerra et al. 2016)

Table	1:	Examples	of	some	mTOR	inhibitors	and	inositol	monophosphatase	inhibitors	experiencing
neuroprotection and the proposed mode of action											

Drug	Model	Mode of action	Major finding	Reference
Rapamycin	Spinal cord injury	Inhibition of	Reduced neuronal damage and	(Sekiguchi
	(SCI) in mice	mTOC1	locomotor impairment through	et al. 2012)
			autophagy activation	
Cystatin C	murine primary	mTOR inhibition	Increased cell survival and had	(Tizon et al.
	cortical neuronal		anti-Aβ amyloidogenetic property	2010)
	cell line and N2a			
	cells			
Lamotrigine	Animal model of	mTOR inhibition	Reduced Ab production by	(Wu et al.
	Alzheimer's	and CREB	reducing the mRNA levels of $\beta$ -	2015)
	disease (AD) in	activation	site ABPP-cleaving enzyme 1	
	AbPP/PS1 mice		(BACE)	
Curcumin	Alzheimer's	Down regulating	Inhibit the generation and	(Wang et al.
	disease in APP/	PI3 K/Akt/	deposition of A <sub>β</sub>	2014)
	PS1 double	mTOR signaling		
	transgenic mice	pathway		
Sodium	Drosophila and	Reduced	Reduced accumulation of	(Williams et
valproate	Zebra fish models	ionositol and IP3	huntingtin and other mutant	al. 2008)
	of Huntington's	levels	proteins	
	disease			
Resveratrol	Rotenone-induced	Heamoxygenase	Reduced apoptosis and oxidative	(Lin et al.
	neurotoxicity in	1 (HO-1)	damage to dopaminergic neurons	2014)
	dopaminergic SH-	dependent		
	SY5Y cells	autophagic flux		
β-Asarone	6-Hydroxy	through	$\beta$ -Asarone improved the	(Zhang et al.
	DopamineInduced	modulation of	behavioral symptoms of rats in the	2016)
	Parkinsonism in	JNK/ Bcl-	open field, rotarod test, initiation	
	rats	2/Beclin-1	time, and stepping time	
		pathway		

### **References:**

Altmann, C., S. Hardt, C. Fischer, J. Heidler, H. Y. Lim, A. Haussler, B. Albuquerque, B. Zimmer, C. Moser, C. Behrends, F. Koentgen, I. Wittig, M. H. H. Schmidt, A. M. Clement, T. Deller and I. Tegeder (2016). Progranulin overexpression in sensory neurons attenuates neuropathic pain in mice: Role of autophagy. Neurobiol Dis 96: 294-311.

Areti, A., P. Komirishetty, M. Akuthota, R. A. Malik and A. Kumar (2017). Melatonin prevents mitochondrial dysfunction and promotes neuroprotection by inducing autophagy during oxaliplatin-evoked peripheral neuropathy. J Pineal Res 62(3).

Berliocchi, L., M. Maiaru, G. P. Varano, R. Russo, M. T. Corasaniti, G. Bagetta and C. Tassorelli (2015). Spinal autophagy is differently modulated in distinct mouse models of neuropathic pain. Mol Pain 11: 3.

Choi, A. M., S. W. Ryter and B. Levine (2013). Autophagy in human health and disease. N Engl J Med 368(19): 1845-1846.

Geng, Y., L. Kohli, B. J. Klocke and K. A. Roth (2010). Chloroquine-induced autophagic vacuole accumulation and cell death in glioma cells is p53 independent. Neuro Oncol 12(5): 473-481.

Glick, D., S. Barth and K. F. Macleod (2010). Autophagy: cellular and molecular mechanisms. J Pathol 221(1): 3-12.

Jung, C. H., S. H. Ro, J. Cao, N. M. Otto and D. H. Kim (2010). mTOR regulation of autophagy. FEBS Lett 584(7): 1287-1295.

Kim, H. J. and H. C. Koh (2021). Chaperonmediated autophagy can regulate diquat-induced apoptosis by inhibiting alpha-synuclein accumulation cooperatively with macroautophagy. Food Chem Toxicol 158: 112706.

Lin, T. K., S. D. Chen, Y. C. Chuang, H. Y. Lin, C. R. Huang, J. H. Chuang, P. W. Wang, S. T. Huang, M. M. Tiao, J. B. Chen and C. W. Liou (2014). Resveratrol partially prevents rotenone-induced neurotoxicity in dopaminergic SH-SY5Y cells through induction of heme oxygenase-1 dependent autophagy. Int J Mol Sci 15(1): 1625-1646.

Longatti, A. and S. A. Tooze (2009). Vesicular

trafficking and autophagosome formation. Cell Death Differ 16(7): 956-965.

Mizushima, N. (2007). Autophagy: process and function. Genes Dev 21(22): 2861-2873.

Mizushima, N., A. Yamamoto, M. Matsui, T. Yoshimori and Y. Ohsumi (2004). In vivo analysis of autophagy in response to nutrient starvation using transgenic mice expressing a fluorescent autophagosome marker. Mol Biol Cell 15(3): 1101-1111.

Mizushima, N., T. Yoshimori and Y. Ohsumi (2011). The role of Atg proteins in autophagosome formation. Annu Rev Cell Dev Biol 27: 107-132.

Oliveira-Abreu, K., F. W. Ferreira-da-Silva, K. S. D. Silva-Alves, N. M. Silva-Dos-Santos, A. C. Cardoso-Teixeira, F. G. D. Amaral, J. Cipolla-Neto and J. H. Leal-Cardoso (2018). Melatonin decreases neuronal excitability in a sub-population of dorsal root ganglion neurons. Brain Res 1692: 1-8.

Ravikumar, B., M. Futter, L. Jahreiss, V. I. Korolchuk, M. Lichtenberg, S. Luo, D. C. Massey, F. M. Menzies, U. Narayanan, M. Renna, M. Jimenez-Sanchez, S. Sarkar, B. Underwood, A. Winslow and D. C. Rubinsztein (2009). Mammalian macroautophagy at a glance. J Cell Sci 122(Pt 11): 1707-1711.

Ravikumar, B., S. Sarkar, J. E. Davies, M. Futter, M. Garcia-Arencibia, Z. W. Green-Thompson, M. Jimenez-Sanchez, V. I. Korolchuk, M. Lichtenberg, S. Luo, D. C. Massey, F. M. Menzies, K. Moreau, U. Narayanan, M. Renna, F. H. Siddiqi, B. R. Underwood, A. R. Winslow and D. C. Rubinsztein (2010). Regulation of mammalian autophagy in physiology and pathophysiology. Physiol Rev 90(4): 1383-1435.

Reichling, D. B. and J. D. Levine (2011). Pain and death: neurodegenerative disease mechanisms in the nociceptor. Ann Neurol 69(1): 13-21.

Schreiber, A. K., C. F. Nones, R. C. Reis, J. G. Chichorro and J. M. Cunha (2015). Diabetic neuropathic pain: Physiopathology and treatment. World J Diabetes 6(3): 432-444.

Sekiguchi, A., H. Kanno, H. Ozawa, S. Yamaya and E. Itoi (2012). Rapamycin promotes autophagy and reduces neural tissue damage and locomotor impairment after spinal cord injury in mice. J Neurotrauma 29(5): 946-956.

Singh, R., L. Kishore and N. Kaur (2014). Diabetic peripheral neuropathy: current perspective and future directions. Pharmacol Res 80: 21-35.

Tan, D. X., L. C. Manchester, E. Esteban-Zubero, Z. Zhou and R. J. Reiter (2015). Melatonin as a Potent and Inducible Endogenous Antioxidant: Synthesis and Metabolism. Molecules 20(10): 18886-18906.

Tanaka, Y., T. Matsuwaki, K. Yamanouchi and M. Nishihara (2013). Exacerbated inflammatory responses related to activated microglia after traumatic brain injury in progranulin-deficient mice. Neuroscience 231: 49-60.

Tizon, B., S. Sahoo, H. Yu, S. Gauthier, A. R. Kumar, P. Mohan, M. Figliola, M. Pawlik, A. Grubb, Y. Uchiyama, U. Bandyopadhyay, A. M. Cuervo, R. A. Nixon and E. Levy (2010). Induction of autophagy by cystatin C: a mechanism that protects murine primary cortical neurons and neuronal cell lines. PLoS One 5(3): e9819.

Towns, R., Y. Kabeya, T. Yoshimori, C. Guo, Y. Shangguan, S. Hong, M. Kaplan, D. J. Klionsky and J. W. Wiley (2005). Sera from patients with type 2 diabetes and neuropathy induce autophagy and colocalization with mitochondria in SY5Y cells. Autophagy 1(3): 163-170.

Wang, C., X. Zhang, Z. Teng, T. Zhang and Y. Li (2014). Downregulation of PI3K/Akt/mTOR signaling pathway in curcumin-induced autophagy in APP/PS1 double transgenic mice. Eur J Pharmacol 740: 312-320.

Williams, A., S. Sarkar, P. Cuddon, E. K. Ttofi, S. Saiki, F. H. Siddiqi, L. Jahreiss, A. Fleming, D. Pask, P. Goldsmith, C. J. O'Kane, R. A. Floto and D. C. Rubinsztein (2008). Novel targets for Huntington's disease in an mTOR-independent autophagy pathway. Nat Chem Biol 4(5): 295-305.

Wong, E. and A. M. Cuervo (2010). Autophagy gone awry in neurodegenerative diseases. Nat Neurosci 13(7): 805-811.

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Wu, H., M. H. Lu, W. Wang, M. Y. Zhang, Q. Q. Zhu, Y. Y. Xia, R. X. Xu, Y. Yang, L. H. Chen and Q. H. Ma (2015). Lamotrigine Reduces beta-Site AbetaPP-Cleaving Enzyme 1 Protein Levels Through Induction of Autophagy. J Alzheimers Dis 46(4): 863-876.

Xilouri, M., O. R. Brekk, N. Landeck, P. M. Pitychoutis, T. Papasilekas, Z. Papadopoulou-Daifoti, D. Kirik and L. Stefanis (2013). Boosting chaperone-mediated autophagy in vivo mitigates alpha-synuclein-induced neurodegeneration. Brain 136(Pt 7): 2130-2146.

Yang, S., C. Xia, S. Li, L. Du, L. Zhang and Y. Hu (2014). Mitochondrial dysfunction driven by the LRRK2-mediated pathway is associated with loss of Purkinje cells and motor coordination deficits in diabetic rat model. Cell Death Dis 5: e1217.

Yerra, V. G., C. Gundu, P. Bachewal and A. Kumar (2016). "Autophagy: The missing link in diabetic neuropathy?" Med Hypotheses 86: 120-128.

Zenker, J., D. Ziegler and R. Chrast (2013). Novel pathogenic pathways in diabetic neuropathy. Trends Neurosci 36(8): 439-449.

Zhang, S., X. H. Gui, L. P. Huang, M. Z. Deng, R. M. Fang, X. H. Ke, Y. P. He, L. Li and Y. Q. Fang (2016). Neuroprotective Effects of beta-Asarone Against 6-Hydroxy Dopamine-Induced Parkinsonism via JNK/Bcl-2/Beclin-1 Pathway. Mol Neurobiol 53(1): 83-94.

Ziegler, D. (2008). Painful diabetic neuropathy: treatment and future aspects. Diabetes Metab Res Rev 24 Suppl 1: S52-57.

Zimmet, P. (2009). Preventing diabetic complications: a primary care perspective. Diabetes Res Clin Pract 84(2): 107-116.